Hypoalgesic effect of *Spirulina platensis* on the sciatic neuropathic pain induced by chronic constriction injury in male rats

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ABSTRACT

Background: It has been revealed that herbal medicines have a palliative effect on pain. In the present study, the hypoalgesic effect of *Spirulina platensis* (microalgae) on the neuropathic pain induced by chronic constriction injury (CCI) was investigated. **Methods**: In the present study, 74 adult male Wistar rats weighing 200-220 grams were used. For inducing neuropathic pain, CCI was performed on the left sciatic nerve. *Spirulina platensis* was intragastrically administered daily for 3 weeks. Mechanical allodynia and thermal hyperalgesia were assessed by Von Frey hairs and plantar test device, respectively. Malondialdehyde (MDA) and total antioxidant capacity (TOC) were detected in the serum using thiobarbituric acid and ferric reducing ability of plasma (FRAP), respectively. **Results**: CCI of the sciatic nerve led to mechanical allodynia and thermal hyperalgesia at three weeks as well as two weeks post surgery. Three weeks of *Spirulina* therapy significantly (P<0.05) decreased paw withdrawal response to mechanical and thermal stimulations, compared to control. FRAP, but not MDA, significantly decreased three weeks after CCI, and *Spirulina* therapy significantly reversed its level towards control. **Conclusion**: Chronic intragastric administration of *Spirulina* platensis alleviates CCI-induced neuropathic pain by modulating oxidative stress through increasing FRAP levels in male rats.

Key words: Chronic constriction injury, Neuropathic pain, Oxidative stress, Rat, Spirulina platensis

INTRODUCTION

Neuropathic pain is one of the major chronic health problems, with allodynia and hyperalgesia being the most common signals of pain¹. Failure in response to standard treatments is a serious challenge in treating these conditions^{2,3}. There are different etiologies for neuropathic pain but one of the most common causes is diabetic neuropathy; in fact, 50-60% of diabetic patients have polyneuropathy⁴. Several complicated and similar mechanisms have been associated with peripheral and central neuropathic pain. These mechanisms include increased release of excitatory neurotransmitters, modulation of descending inhibitory pathways, inflammatory reactions and oxidative stress⁵⁻⁷, all of which may change with time thereby increasing complications⁸. With regard to these complications, all of the approaches presented above have not shown sufficient evidence of effective neuropathic pain treatment^{3,9}. Furthermore, it should be noted that in addition to the varied side effects, standard pharmacologic medications also have serious interactions with each other, which must be considered. Therefore, alternative methods of suppressing and treating neuropathic pain are necessary. Plants have many valuable constituents that enable them to improve cellular function in healthy and

diseased states. Plants and medicinal herbs have been used for a long time ago throughout the world for promoting health and as therapeutic agents in various disorders, such as cancer¹⁰, neuropathy^{11,12}, and Parkinson's disease¹³. Among plants, Spirulina platensis is a marine microalgae which belongs to the class of cyanobacteria¹⁴, known for containing rich resources of carotenoids, trace elements, and vitamins¹⁵. Spirulina has many positive biological features, including anti-inflammatory and antioxidant^{16,17}, antimicrobial¹⁸, antitumoral¹⁹ and immunomodulatory properties. It was reported that Spirulina alleviates proinflammatory cytokine TNFalpha in the brain tissues of aged rats¹⁷. The antioxidant property of Spirulina is ascribed to its Cphycocyanin component^{20,21}, which is a potent antioxidant.

There are several animal studies demonstrating the protective effects of *Spirulina* against environmental chemotoxicity and pharmacologic-induced oxidative stress²². For example, it has been reported that *Spirulina* reduces neuronal injury due to oxidative stress and ischemia^{23,24}. It has also been reported that *Spirulina* protects different organs against toxicity of heavy metals and pharmacologic agents^{25–27}. Indeed,

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Spirulina completely blocked MPTP (1-methyl-4phenyl-1, 2, 3, 6- tetrahydropyridine)-induced oxidative stress, partially reduced MPTP-induced Parkinson's disease symptoms in mice, and improved nigrostriatal dopamine pathway injury (caused by 6hydroxydopamine (6-OHDA)) in rats^{23,28}. In light of these effects, it seems that the protective effects of Spirulina are, at least, due to its antioxidant properties, although other mechanisms may also play a role. Given the aforementioned impact of the role of oxidative stress and inflammation in peripheral neuropathic pain⁶, and the antioxidative properties of Spirulina microalgae, the aim of this study was to evaluate whether orally administered Spirulina could suppress neuropathic pain induced by chronic constriction injury (CCI) in rats. To our understanding, no other study has examined the effects of Spirulina therapy on CCI-induced neuropathic pain thus far.

METHODS

Animals

In this study, adult male Wistar rats weighing 200-220 grams were used. Animals were obtained from the animal house of Semnan University of Medical Sciences. The rats were kept in an environment where humidity, temperature, and light-dark cycle were controlled. Food and water were sufficiently available to the animals. All experimental procedures were performed, as approved by the local ethical committees on animal research of the Semnan University of Medical Sciences, under permit number 1394.217.

Drug

Spirulina platensis microalgae were provided from Qeshm Microalgae Biorefinery Company (Qeshm Island, Iran). *Spirulina* was in a powder form.Spirulina was dissolved in physiologic saline at concentrations of 200, 400, and 800 mg/kg and was administered intragastrically to the rats in a volume of 2 ml by a gavage tube. The dosage of *Spirulina platensis* evaluated in our study were: 0, 200, 400, and 800 mg/kg.

Induction of neuropathy

Induction of neuropathy was performed through the method described by Bennett and Xie²⁹. Animals were anesthetized using intraperitoneal injection of ketamine (80 mg/kg) and xylazine (10 mg/kg) mixture. Using a surgical blade, a 2-cm incision was made on the left thigh and the sciatic nerve was exposed and then released from the surrounding tissues. Using catgut chromic sutures 4-0,4 loose ligations were made around the common sciatic nerve at

1-mm intervals. At the end, the incisures sites (muscle and skin) were separately sutured with silk 4-0. In the sham group, after exposing the sciatic nerve only, muscle and skin were sutured.

Experimental protocol and groups

In this study, 74 male rats were used in two experiments. In the first experiment, 37 rats were randomly assigned to 6 groups (Intact, Sham, Neuropathy, Neuropathy with Spirulina (200 mg/kg), Neuropathy with Spirulina (400 mg/kg), and Neuropathy with Spirulina (800 mg/kg)) for behavioral and oxidative stress tests at 14 days post surgery. In the second experiment, another 37 rats were assigned to the same groups and tested at 21 days post surgery. Saline or Spirulina gavages were initiated on the post surgery day and continued on to 14 and 21 days after surgery in the first and second experiments, respectively. Spirulina was dissolved in physiologic saline and was intragastrically administered in a volume of 2 ml by a gavage tube. Four rats were paralyzed and thus excluded from the study. The study design is demonstrated in Table 1.

Assessment of pain behavior of the animals

Pain response was evaluated using the following tests:

Mechanical allodynia

Paw withdrawal response was detected using the von Frey's hairs (Stoelting, Wood Dale, IL, and USA) which were described by Ren *et al.*³⁰. Von Frey's hairs are calibrated filaments that, depending on their thickness, apply a certain force (in grams) to the surface to which they are pressed. The filaments were applied with ascending style on the dorsal surface of the injured hind paws. The experiment began with a low force and if there was no response, a greater force was applied. If the animal withdrew its foot three times per five stimuli delivered, that force was considered as a response. A stimulus equivalent to 60 g was considered as the cut-off force.

Thermal hyperalgesia

The animals were placed in the plantar test device and after habituation, infrared radiation was applied to the plantar surface of the injured paw with an intensity of sixty. Infrared radiation was displayed every five minutes in three rounds. Withdrawal latency to each thermal stimulus was recorded and the average of three rounds was considered as response. Sixty seconds was set as cut-off point of the test²⁹.

1 07 0	Table 1: Design of the study				
Experiment 1 Day 0 Days 1-13 Day 14	Groups	Surgery	Gavages	Behavioral, oxidative stress tests	
Experiment 1 Day 0 Days 1-15 Day 14	Experiment 1	Day 0	Days 1-13	Day 14	
Experiment 2 Day 0 Days 1-20 Day 21	Experiment 2	Day 0	Days 1-20	Day 21	

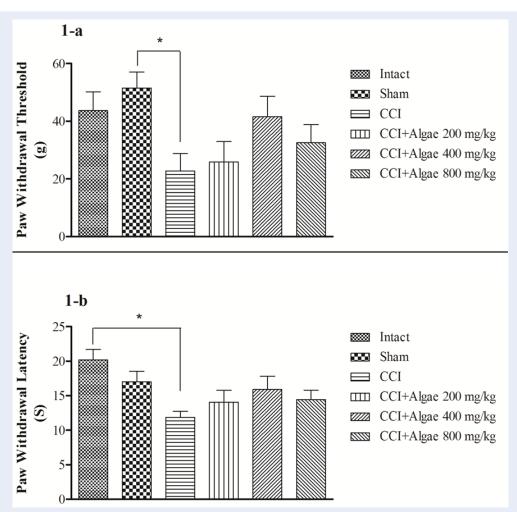


Figure 1: Effect of *Spirulina Platensis* microalgae on the chronic constriction injury-induced neuropathic pain at 14 days post lesion in male rats. Chronic constriction injury of sciatic nerve led to significant tactile allodynia (A) and thermal hyperalgesia (B). Administration of Spirulina did not change mechanical allodynia and thermal hyperalgesia at 2 weeks post injury. All data are expressed as mean \pm S.E.M of measured variable. N=7-9. **P*<0.05

Assessment of oxidative stress reaction

Oxidative stress was evaluated using malondialdehyde (MDA) and ferric reducing ability of plasma (FRAP) assay of the rats' blood.

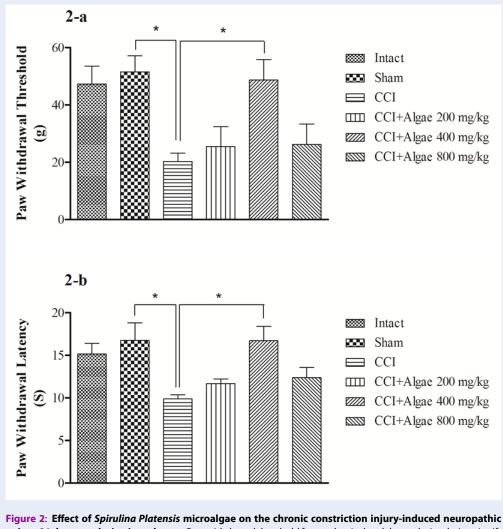
Blood sampling

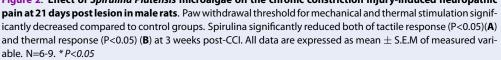
Blood samples were prepared from the rats' hearts while they were anesthetized. Using a centrifuge with

2000 rpm for 10 min, the serum was removed from the blood and kept at -80° C until use in the MDA and FRAP assays.

MDA assay

The MDA was assayed in rat serum. Thiobarbituric acid method was used to measure lipid peroxidation³¹. Peroxidation of unsaturated fatty acids increases through oxidative stress, and various alde-





hydes (including MDA) are produced. MDA (as a marker of oxidative stress) reacts with thiobarbituric acid in acidic regions at high temperatures. The maximum absorption was evaluated at 535 nm using a spectrophotometer.

FRAP assay

For detecting FRAP, measurement of total antioxidant capacity (TAC) in the rat serum was used. The ability of the serum to reduce Fe³⁺ ions to Fe²⁺ is the basis of this method and the output is a blue complex solution which shows the maximum light absorption at 593 nm wavelength ³².

Statistical analysis

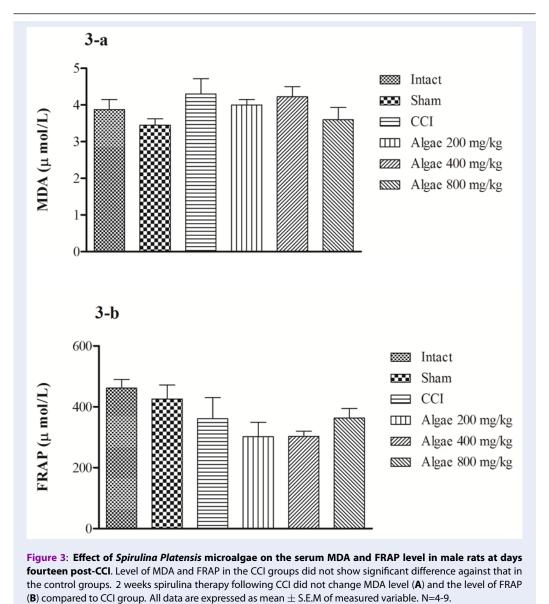
One-way analysis of variance (ANOVA) and Tukey's post-hoc test were used for analyzing the data. All data were expressed as mean \pm SEM of the detected variables; P<0.05 was considered significant.

RESULTS

Our results revealed that *Spirulina* can suppress neuropathic pain induced by CCI. The results were divided in two parts- behavioral and biochemical.

Behavioral results

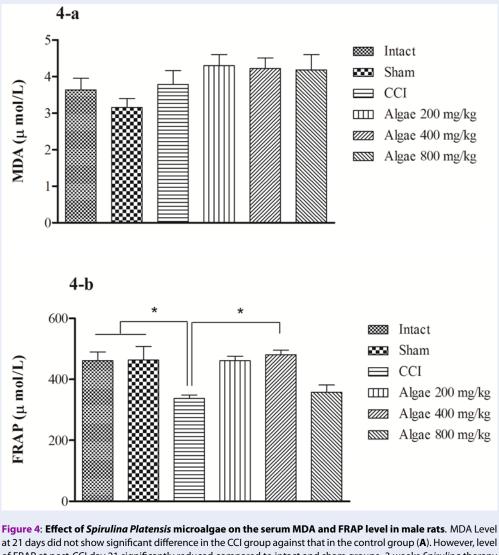
We evaluate the effects of *Spirulina platensis* on neuropathic pain in rats. We found that *Spirulina* therapy

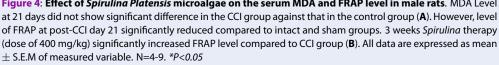


significantly reversed CCI-induced neuropathic pain, relative to control group, after 21 days of treatment. From experiment 1, mechanical allodynia and thermal hyperalgesia were significantly reduced in CCI groups compared to intact and sham groups, respectively (**Figure 1A, B**). However, neuropathic pain was not improved by *Spirulina* therapy. Paw withdrawal responses (mechanical and thermal) were not significantly different in the treatment groups, compared to CCI group, at 14 days post CCI (**Figure 1A, B**). In Experiment 2, mechanical allodynia and ther-

mal hyperalgesia were significantly reduced in CCI groups, compared to control/sham groups (**Figure 2A, B**). *Spirulina* significantly improved

neuropathic pain at 21 days post therapy. Our results showed that *Spirulina* therapy significantly decreased mechanical allodynia [P<0.05, F (5, 38) =5.07] and thermal hyperalgesia [P<0.05, F (5, 34) =4.45], compared to CCI group (**Figure 2A, B**). The pain reducing effect of *Spirulina* was observed with moderate dose (400 mg/kg), as tested in the present study. The effect of *Spirulina* on pain behavior showed an inverse U pattern such that a reducing effect on neuropathic pain was observed with a dose of 400 mg/kg but not with 200 or 800 mg/kg.





Oxidative stress results

Using MDA and FRAP as oxidative markers, we studied the oxidative stress alteration during *Spirulina* therapy in a CCI-induced neuropathic pain situation. In both experiments, the MDA level was not significantly different between the groups (**Figure 3A** and **Figure 4A**). Like MDA levels, FRAP levels in Experiment 1 was not significantly different between that of the control, CCI, or treatment groups (**Figure 3B**). On the other hand, FRAP was significantly reduced compared to intact and sham groups in experiment 2 (**Figure 4B**). Treatment with *Spirulina* (400 mg/kg) led to a significant increase [P<0.05, F (5, 33) =4.5] of FRAP levels at 21 days post CCI. However, FRAP measurement did not show improvement at 14 days post CCI.

DISCUSSION

Neuropathic pain is a severe debilitating problem following somatosensory injury. Several reports have revealed the efficacy of medicinal herbs in treating neuropathic pain ^{33–35}. In the present study, we evaluated the efficiency of *Spirulina platensis* microalgae on alleviating neuropathic pain induced by CCI of sciatic nerve in male rats. To our knowledge, this is the first study to study the effects of *Spirulina platensis* on sciatic neuropathic pain treatment.

Hypoalgesic effects

Our results revealed that CCI leads to mechanical allodynia and thermal hyperalgesia, consistent with findings from our previous study⁶. Spirulina microalgae significantly reduced CCI-induced neuropathic pain. Intragastric administration of Spirulina powder significantly increased paw withdrawal threshold and paw withdrawal latency in neuropathic animals at 21 days after induction of neuropathy (Figure 2). Nisha Patro et al. reported that oral use of Spirulina alleviates arthritis pain through suppressing inflammatory processes and microglial activation in the spinal cord³⁶. Previously, we reported that inflammatory mediator release due to CCI is one of the main mechanisms leading to neuropathic pain⁶. Consistent with the studies which suggest an anti inflammatory property for Spirulina and its content Cphycocyanin^{17,37,38}, it is possible that in the present study Spirulina led to analgesia through suppressing inflammatory mediators. In the present study, the hypoalgesic effect of Spirulina was observed when using a moderate dose of Spirulina (400 mg/kg). Consistent with our results, Patro et al. showed that a dose of 400 mg/kg was effective in their behavioral (sciatic function index) and molecular (downregulation of IbA1 as an index of microglial activation at both of ventral and dorsal horns of the spinal cord) experiments compared to higher doses³⁶. In the present study, the doses of 200 and 800 mg/kg did not show any positive effect on reducing neuropathic pain. Spirulina administration showed an inverse U effect on behavioral tests.

Antioxidant effects

The results of this study showed that the antioxidant properties of phytochemicals alleviate neuropathic pain. Our results showed that *Spirulina* has an antioxidant capacity. We investigated the antioxidant activity of *Spirulina* by performing FRAP and MDA assays on the serum of chronic constriction injured rats.

In our study, MDA as an oxidative stress marker did not change between control and treatment groups. Several *in vitro* and *in vivo* studies revealed the antioxidant properties of *Spirulina* through decrease of MDA levels^{6,27,39}. However, the present results are consistent with our previous results⁶; MDA levels were not significantly different between the control and neuropathy groups. It is may be that in the present model, changes in the MDA levels makes longer period of time. However, more research on this topic needs to be undertaken. Another possible explanation for this observation is that other antioxidant biomarkers may have changed.

Our results revealed that FRAP levels were significantly reduced in neuropathic rats but prominently increased following Spirulina therapy (Figure 4). It has been reported that pre-treatment with Spirulina reduces motor disturbances following spinal cord injury in rats⁴⁰. Moreover, it was reported that Spirulina improves nigrostriatal system injury in the rats²⁸. Consistent with our results, there have been several reports about the role of Spirulina in the increasing the level of TOC. It has been shown that drug and chemical-induced histopathological damages were alleviated by Spirulina therapy in rats^{22,26}. Khan and his colleagues, in 2005, showed that Spirulina has a protective effect on doxorubicin-induced damage to heart through normalizing antioxidant enzymes⁴¹. Thaakur et al. showed that vascular ischemic injury was suppressed by Spirulina and that this effect seemed to be due to the antioxidant activity of Spirulina²⁴.

In addition to animal studies, the efficacy of Spirulina as an antioxidant has been demonstrated in clinical studies. Orally administered Spirulina (daily dose of 8 g for 4 weeks) led to a significant increase of TOC, which occurred in parallel to prolongation of fatigue during physical activity in male human subjects⁴². Spirulina is a rich source of Cphycocyanin^{22,43}. In fact, various studies have reported that C-phycocyanin has an antioxidative ability^{44,45}. It was reported that orally administration of C-phycocyanin protected the rat hippocampus neurons against kainic acid-induced injury through antioxidant activity⁴⁶. Romay et al., in 1998, showed the ability of phycocyanin in scavenging hydroxyl and alkoxyl radicals⁴⁷. It has been reported that the antioxidant activity of phycocyanin provides anti-inflammatory effects in animal models, such as arachidonic acid-induced ear edema in mice and carageenan-induced paw edema in rats³⁸.

Considering the reports mentioned above, it seems that the suppressive effect of *Spirulina* on CCIinduced neuropathic pain is due to its antioxidant ability through enhancement of plasma TOC or through inhibiting inflammatory processes. In the future, it is important to evaluate in further detail the inflammatory mediators, such as TNF-alpha, and antioxidant biomarkers, such as superoxide dismutase and glutathione. This was one of the limitations of the present study; *i.e.* we did not evaluate additional mediators and biomarkers due to expenses.

CONCLUSION

The most obvious finding to emerge from this study is that *Spirulina* microalgae can reduce neuropathic pain induced by chronic constriction injury in male rats. This effect is caused by the antioxidant activity of *Spirulina* through increasing FRAP levels.

COMPETING INTERESTS

All authors have nothing to disclose and have no financial or commercial conflict of interests in the present study.

AUTHORS' CONTRIBUTIONS

Dr. Ghanbari and Mr Safakhah carried out the design and coordinated the study, participated in most of the experiments. Dr. Bandegi, Mr Moradi kor and Miss Tamimi carried out all the experiments. Dr. Ghanbari, Dr Bandegi and Mr Safakhah carried out the Analysis of data. Dr. Ghanbari, Mr Safakhah and Miss Tamimi participated in manuscript preparation. All authors have read and approved the content of the manuscript.

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ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Ethics approval for this study was obtained from the Institutional Review Board of Semnan University of Medical Sciences, in order to the Helsinki declaration (No 1394; 217)

AVAILABILITY OF DATA AND MATERIALS

Authors will provide if requested.

FUNDING

Semnan University of Medical Sciences, Semnan, Iran.

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